

Review

Oxidative Modification of Brain Proteins in Alzheimer's Disease: Perspective on Future Studies Based on Results of Redox Proteomics Studies

Rukhsana Sultana^{a,*} and D. Allan Butterfield^{1a,b,c,*}

^a*Department of Chemistry, University of Kentucky, Lexington, KY, USA*

^b*Center of Membrane Sciences, University of Kentucky, Lexington, KY, USA*

^c*Sanders-Brown Center on Aging, University of Kentucky, Lexington, KY, USA*

Abstract. Aging is the major risk factor associated with neurodegenerative diseases, including Alzheimer's disease (AD). Until now no clear understanding of the mechanisms of initiation and progression of this dementing disorder exists. Based on the studies that have been conducted so far amyloid β -peptide (A β), a protein found in senile plaques, one of the key pathological hallmarks of AD, has been reported to be critical in the pathogenesis of AD. Studies from our laboratory and others showed that A β can induce oxidative stress, which leads to oxidative modification of biomolecules, thereby diminishing the normal functions of neuronal cells and eventually leading to loss of neurons and AD. In this review paper, we summarize evidence of oxidative stress in brains of AD and mild cognitive impairment patients, as well as the results from redox proteomics studies. The investigations have provided insights into the downstream effects of oxidative modification of key brain proteins in the pathogenesis of AD. Based on these redox proteomics results, we suggest future areas of research that could be considered to better understand this devastating dementing disorder.

Keywords: Alzheimer's disease, lipid peroxidation, mild cognitive impairment, oxidative stress, protein carbonylation, protein nitration, redox proteomics

INTRODUCTION

We appreciate being invited to share perspectives on our prior publications of highly cited papers

*Correspondence to: Prof. D. Allan Butterfield, Department of Chemistry, Center of Membrane Sciences, and Sanders-Brown Center on Aging, University of Kentucky, Lexington, KY 40506-0055, USA. Tel.: +1 859 257 3184; Fax: +1 859 257 5876; E-mail: dabcs@uky.edu or Prof. Rukhsana Sultana, Department of Chemistry, University of Kentucky, Lexington, KY 40506-0055, USA. Tel.: +1 859 257 3615; Fax: +1 859 257 5876; E-mail: rsult2@uky.edu.

on Alzheimer's disease (AD), many of which have dealt with oxidative stress and redox proteomics analysis in AD and amnesic mild cognitive impairment (MCI), and on some future studies these papers suggest. Accordingly, this review paper summarizes evidence of oxidative stress in AD and MCI brain and results using redox proteomics, followed by our view of some future directions in AD research. Aging is the major risk factor associated with neurodegenerative diseases, including AD, which is histopathologically characterized by the presence of senile plaques (SP), neurofibrillary tangles

(NFTs), and loss of synapses [1]. However, until now the exact mechanism(s) of AD progression or pathogenesis largely remain unknown. Studies involving familial AD suggested that the mutations of *presenilin-1*, *presenilin-2*, and *amyloid- β protein precursor (A β PP)* genes cause familial AD (FAD), and implicate amyloid β -peptide (A β) as the underlying cause for the onset of pathology, clinical presentation, and biochemical alterations in this devastating disease. In addition, mutation in other genes, such as *apolipoprotein E* allele 4 (*APOE 4*), *endothelial nitric oxide synthase-3*, *phosphatidylinositol-binding clathrin assembly protein (PICALM)*, *clusterin (CLU)*, also called *apolipoprotein J*, and *α 2-macroglobulin*, have been suggested as risk factors for AD [2–5].

The main component of the SP is a 40–42 amino acid peptide, A β , generated by the proteolytic cleavage of A β PP by the action of β - and γ -secretases [6]. The A β -peptide exists in both soluble and insoluble forms, and has been shown to be toxic. The toxicity induced by A β has been associated with the single methionine (Met) residue present at 35 position in A β peptide [7, 8]. Met can undergo one-electron oxidation to form sulfuranyl or hydroxysulfuranyl radical cations, which can abstract allylic hydrogen

atoms from phospholipid acyl chains, thereby initiating the lipid peroxidation via chain reaction processes and consequently in the generation of highly reactive products such as 4-hydroxy-2-*trans* nonenal (HNE) and acrolein [7, 9]. Since the plasma membrane and organelle membranes have both lipid and protein components, the generation of reactive products like HNE by lipid peroxidation makes the membrane proteins in the membrane highly susceptible to oxidative modification via Michael addition, which affects the protein structure and eventually impairs cellular function as reported in AD [10, 11] (Fig. 1). Studies from our laboratory and others showed that lack of Met leads to significantly diminished oxidative stress [12–14].

The importance of A β in inducing oxidative stress and being a key player in AD pathogenesis is supported by the studies involving Down syndrome, individuals characterized by a trisomy of chromosome 21. The extra copy of A β PP gene in these individuals leads to increased levels of A β , and correlated with increase oxidative stress and AD-like pathology if they live long enough [15, 16]. However, until now it is not clear what the first step is that leads to increasing load of A β . Individuals with AD have elevated

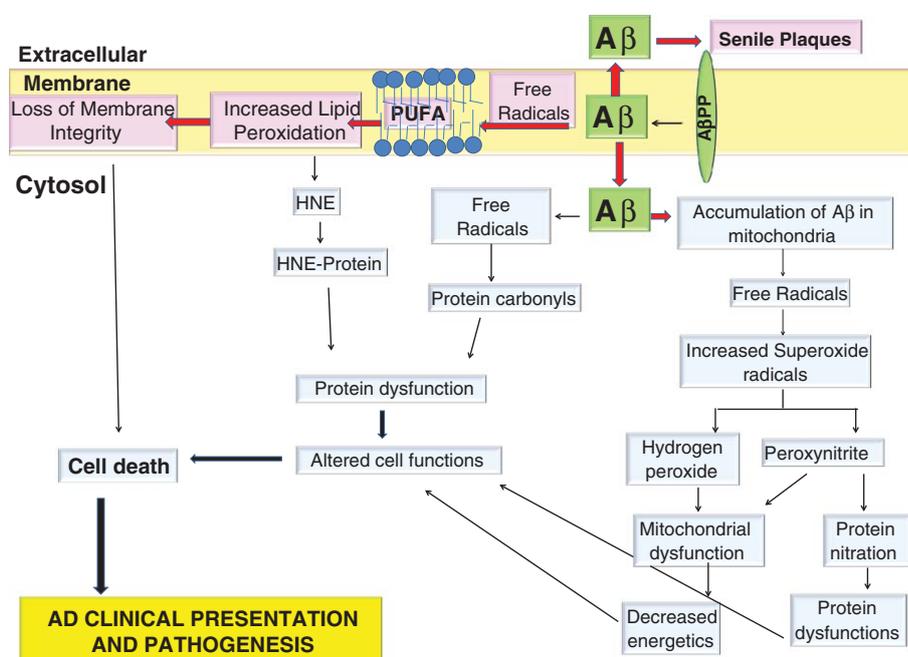


Fig. 1. Amyloid- β (A β) peptide generated by the proteolytic cleavage of amyloid- β protein precursor (A β PP) is important in inducing oxidative and nitrosative damage in AD and is key in the progression of this dementing disorder. Lipid peroxidation induced by bilayer-soluble oligomeric A β _{1–42} and mitochondrial accumulation of this neurotoxic peptide lead to a cascade of events schematically depicted here. Other cellular changes are not indicated, but may include oxidation of nuclear and mitochondrial DNA.

levels of cerebral A β and increased markers of oxidative stress such as protein carbonylation, protein nitration, HNE, acrolein, 8-hydroxy guanosine, and advanced glycation end products. [17–28]. A β -induced oxidative stress is further supported by a number of *in vitro* and *in vivo* studies [29–33]. Further, elevated levels of oxidative stress have also been found in brains from subjects with MCI, arguably the earliest form of AD, and early AD [21, 26, 29, 34–36]. Individuals with preclinical AD (PCAD) have significant levels of A β deposition, but no significant increase in oxidative stress compared to age-matched control, which is consistent with the recent finding that small oligomers of A β are highly toxic in these individuals. Indeed, the oligomeric form of A β is of lower concentration in brains of PCAD subjects compared to AD and MCI [37, 38]. A β ₁₋₄₂ has been shown to aggregate more quickly than A β ₁₋₄₀ and is proposed to play a central role in AD pathogenesis. The evidence of A β ₁₋₄₂ being involved in AD pathogenesis is largely derived from the observation that FAD cases have increased A β load and increased oxidative stress [39].

OXIDATIVELY-MODIFIED PROTEINS

Early approaches used to gain insight into the specific protein targets of oxidative modification involved immunoprecipitation techniques, in which a protein of interest was immunoprecipitated and then probed with the antibody of the type of modification that was to be tested. This technique is labor intensive, time consuming, and additionally requires the availability of the antibody of the protein that needed to be tested. To achieve the goal of identification of multiple oxidatively-modified brain proteins simultaneously, our laboratory pioneered a technique called redox proteomics [40–44]. One manifestation of redox proteomics couples two-dimensional gel electrophoresis (2D) with isoelectrofocusing (IEF) to separate the large number of brain proteins followed by protein transfer to a 2D Western blot on which proteins are probed for protein carbonyls, 3-NT, or bound HNE [41, 43, 45–47]. Sophisticated imaging analysis, coupled with trypsin digestion and use of software, identified spots of interest, and mass spectrometry allowed interrogation of protein databases that led to the identification of a large number of targets of oxidation.

PROTEIN OXIDATION: CARBONYLATION AND NITRATION

Beta-actin and creatine kinase BB have been identified as specifically oxidized proteins in AD brain using 2D electrophoresis and 2D Western blots [48]. These techniques form the basis of the methodology needed to further examine the role of oxidative modifications of specific brain proteins in AD pathogenesis and have led to the development and use of redox proteomic [44] techniques to identify carbonylated brain proteins in AD [41, 42, 49]. 2D gel electrophoresis coupled with mass spectrometry [44, 45] have allowed the discovery of increased carbonylation creatine kinase BB, glutamine synthase, ubiquitin carboxy-terminal hydrolase L-1 (UCH L-1), dihydropyrimidinase-related protein 2 (DRP-2), α -enolase and heat shock cognate 71 in AD inferior parietal lobule (IPL) compared to age-matched controls [41, 42, 50]. Subsequent studies of AD hippocampus demonstrate specific carbonylation of peptidyl prolyl cis-trans isomerase (Pin1), phosphoglycerate mutase 1, UCH L-1, DRP-2, carbonic anhydrase II, triose phosphate isomerase (TPI), α -enolase, and γ -SNAP compared to age-matched controls [45]. Consistent with the notion that oxidative modification of proteins leads to dysfunction of normal cellular processes in AD, the activities of Pin1, enolase, and carbonic anhydrase II were significantly lower in AD hippocampus compared to matched tissue samples from control subjects [45]. Alterations in enzymatic function in these systems could contribute to pathogenesis of AD through inhibition of cellular degradation machinery, alteration of protein conformation, decreased cerebrospinal fluid production, and impairment in cellular metabolic processes.

Others [51] using redox proteomics showed significant decreased protein carbonyls in malate dehydrogenase 1 (MDH), glutamate dehydrogenase, 14-3-3 protein zeta/delta, aldolases A and C, and increased oxidation of carbonic anhydrase 1. The sample processing in this study did not use detergents and may have led to identification of fewer oxidized proteins than that seen in other studies as a result of decreased exposure of protein carbonyls. More recent studies identified DJ-1 as a carbonylated protein in the frontal cortex of AD patients [52]. In the IPL of FAD subjects, increased carbonylation of UCH-L1, γ -enolase, actin, and dimethylarginine dimethylaminohydrolase 1 (DMDMAH-1) have been reported [35]. Others also reported oxidation and accumulation of proteins like UCH L1, ATP synthase, and Cu,Zn-superoxide

dismutase in AD brain [49, 53, 54], confirming our prior results.

Many of the proteins that are oxidatively modified in AD brain have roles in energy metabolism. This observation may contribute to the results of PET studies that demonstrate decreased glucose utilization in AD brain. The extent of oxidatively-modified brain proteins by proteomics correlates well with AD pathology, including both SP and NFT burden [41–43, 45]. Further, the identification of common targets of protein carbonylation between FAD, sporadic AD, and MCI is consistent with the idea that increased oxidative stress is invariable in respect to cause (genetic versus sporadic) or stage (amnesic MCI versus dementia) in the pathogenesis of AD [29].

Brains from subjects with MCI also demonstrates increased levels of protein carbonyls [26, 45, 55]. Redox proteomics studies in MCI hippocampus led to the identification of α -enolase, glutamine synthetase, pyruvate kinase M2, and Pin1 as specifically carbonylated proteins, recapitulating many of the findings seen in fulminate AD brain tissue [34]. Recent reports identified oxidatively modified carbonic anhydrase II, heat shock protein 70, mitogen activated protein kinase I, and syntaxin binding protein I in MCI indexed by elevated protein carbonyls [56].

The redox proteomic studies of brain from subjects with AD and MCI identified proteins such as enolase, Pin1, and glutamine synthetase as targets of carbonylation common to both AD and MCI. The functions of these proteins are important not only in regulating energy metabolism, but have also been linked to tau hyperphosphorylation, alterations in protein conformation, A β PP processing, and glutamate regulation, all of which are thought to be relevant to neurodegenerative processes in AD [56]. Of interest, not all brain proteins that appear to be targets of protein carbonylation at an early stage (MCI) appear in advanced stage (AD). This suggests that the specific targets of oxidative stress vary with stage of disease and represent a specific rather than non-selective injurious process. Such findings may have implications for the use of specific disease modifying treatments in relation to stage of disease. It is possible that oxidatively induced alterations in specific cellular pathways contribute specifically to the disease process early on, altering transcription and translational mechanisms that may further damage neurons. Such transcriptional and translational alterations can further reduce the specific substrates for carbonylation, exacerbating the loss of function caused by oxidation early in the disease process. As neuronal injury ensues, the effects of oxidative

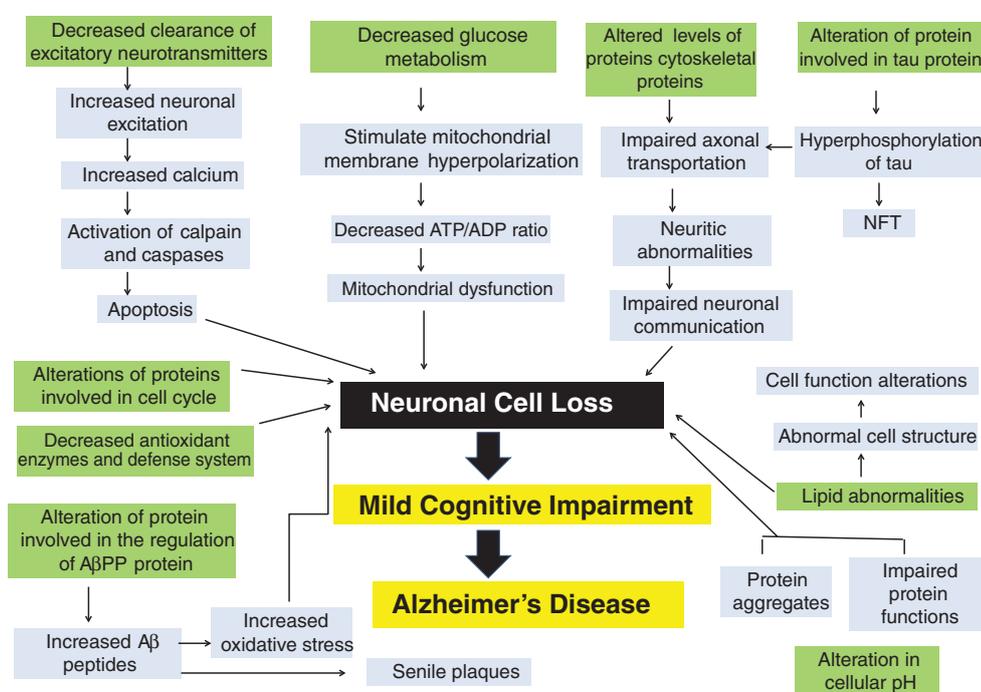


Fig. 2. The oxidative modification of proteins affects a number of pathways in AD and amnesic MCI, arguably the earliest form of AD. The fact that the same pathways are affected in MCI as in AD suggest that alterations in these pathways could be involved in the progression of amnesic MCI to AD. Depicted in this figure are some of these pathways implicated by oxidatively-modified proteins identified by redox proteomics.

damage become more widespread and non-selective as part of the end-stage process of AD.

Figure 2 presents potential pathways affected by protein oxidation in AD and amnesic MCI revealed by redox proteomics. Overlap of pathways in these two conditions involved in energy metabolism, neurotransmitter function, neuritic abnormalities, cell cycle, tau phosphorylation, A β production, pH regulation, and antioxidant system are consistent with the notion that these pathways could be involved in the progression of amnesic MCI (with memory loss) to AD (with dementia).

PROTEIN NITRATION IN AD AND MCI

Redox proteomics studies have identified a large number of proteins that have specifically nitrated Tyr residues in AD hippocampus and IPL compared to control brain, including α - and γ -enolase, lactate dehydrogenase, neuropolypeptide h3, TPI, and α -actin in AD IPL [43], and α -enolase, glyceraldehyde-3-phosphate dehydrogenase (GAPDH), ATP synthase α -chain, carbonic anhydrase-II, and voltage-dependent anion channel protein in AD hippocampus [19]. These nitrated proteins are involved in various cellular functions such as energy metabolism, structural maintenance, pH regulation, and mitochondrial function. Oxidative modification (i.e., nitration, carbonylation, etc.) may alter protein functionality [19]. Our redox proteomics finding of excess nitration of TPI was recently confirmed by Guix et al. [57] in hippocampus and frontal cortex of AD subjects, suggesting a link among decreased glucose metabolism via an impaired glycolytic pathway, nitrosylation of TPI, and the formation of A β and paired helical filaments. However, it is not clear why, in spite of oxidative modification, its activity remains unchanged in AD brain. In contrast, Reyes et al. demonstrated nitration of Tyr 18 followed by Tyr 29 of tau, which is mostly associated with or in close proximity to amyloid plaques [58]. Hence, nitration of proteins may reflect underlying posttranslational modification of proteins in AD.

Consistent with this notion, increased levels of 3-NT in MCI hippocampus and IPL using immunohistochemistry were reported [59]. There is also evidence for AD-specific nitration of MDH, α -enolase, glucose regulated protein precursor, aldolase, glutathione-S-transferase Mu, multidrug resistant protein-3, and 14-3-3 protein γ in MCI IPL [46]. In MCI hippocampus, α -enolase, MDH, peroxiredoxin 6 (PR VI), DRP-2, fascin 1, and heat shock protein A8

were identified as specifically nitrated compared to age-matched controls [46]. These redox proteomics-identified nitrated proteins in MCI are involved in the regulation of a number of important cellular functions including: energy metabolism, cellular signaling, antioxidant, and detoxification, in addition to regulating structural functions of brain cells. The identification of some of the brain protein targets of nitration in common between amnesic MCI and AD is consistent with the notion that these brain proteins might contribute to the progression of and increased synapse and functional loss in AD [19, 43, 46]. We showed increased nitration of p53 protein in MCI IPL compared to age-matched control and suggested that the oxidation of p53 may be involved in neuronal loss [60].

A recent study by Reiderer et al. [61] reported S-nitrosyl-cysteine modification of DRP-2, α -internexin, glutamate dehydrogenase 1, α -enolase, GFAP, MDH, ProSAAS precursor protein, proopiomelanocortin, proenkephalin, and septin in the entorhinal cortex of AD, and suggested that A β activation of glial cells surrounding the SP might have led to increased nitrosylation of GFAP contributing to the pathogenesis of AD. Protein disulfide isomerase, an enzyme that catalyzes thiol-disulphide exchange, has been reported to be S-nitrosylated in AD brain [62]. Increased nitrosylation and decreased activity of this protein in AD may lead to alteration in its ability to facilitate disulfide bond formation and rearrangement reactions, increased accumulation of polyubiquitinated proteins, and activation of the endoplasmic reticulum-resident unfolded protein response. Recently Cho and colleagues [63] reported increased levels of S-nitrosylation of dynamin-related protein 1 in brains of subjects with AD and suggested that S-nitrosylation of this protein may trigger mitochondrial fission, consequently adding to known mitochondrial damage in AD, which could contribute to synapse loss and neuronal damage in this disorder.

Taken together, these studies suggest that oxidation of proteins is an integral part of the progression and pathophysiology of AD [64]. The appearance of common targets of oxidation of proteins between MCI and AD implies their important roles in loss of cellular energetics, alterations in neurotransmission and cell signaling pathways, as well as SP and NFT formation (Table 2).

Enolase, an oxidatively-modified protein in AD and MCI brain, is important for regulating glucose metabolism. However, a number of recent studies showed that enolase also plays important roles in cell

signaling, A β clearance, and activation of cell survival pathways [65]. This result demonstrated that oxidative dysfunction of one protein may alter several cellular pathways implicated in the pathogenesis of AD. This point is further illustrated by GAPDH, which is also selectively oxidized in AD [19]. GAPDH is a key enzyme in the glycolytic pathway; however, recent studies suggest that it may also play key roles in transcription regulation, cell signaling, and vesicular transportation [66, 67], in addition to binding to other small molecules such as nitric oxide, glutathione, and tumor necrosis factor- α [12, 68, 69]. GAPDH also interacts with A β PP [70]. Hence, oxidative dysfunction of enolase and GAPDH can lead to multiple changes consistent with pathology, biochemistry, and clinical presentations of AD and MCI. Modulation of the cellular pathways altered by the selective oxidation of both GAPDH and enolase could prove to be fertile ground for the development of novel therapeutic agents for AD [12, 65].

Another protein that exemplifies how oxidative modification of one protein can significantly affect function of AD-relevant pathways and be an important therapeutic target is Pin1. By its isomerization of proline on the carboxyl side of phosphorylated serine or threonine residues of target proteins, the regulatory protein Pin1 has been shown to play an important role in tau phosphorylation/dephosphorylation, A β PP regulation and processing, and synapse loss [71–73]. Hence, oxidatively dysfunctional Pin1 conceivably could be related to the major pathologies of AD: SP, NFT, and synapse loss [45]. Consequently, targeting Pin1 to treat AD might be a promising approach to treat or delay the onset of AD pathogenesis.

In addition, antioxidant enzymes were also found to be oxidized in common between AD and MCI brain, and loss of their function would have severe effects on cell survival [74]. As discussed further below, redox proteomic discoveries suggest several possible therapeutic strategies that may modulate AD progression and pathogenesis: 1) upregulate the endogenous levels of key oxidatively-modified proteins; 2) restore function in key oxidatively-modified proteins; or 3) augment cellular antioxidant systems.

HNE ADDUCTED BRAIN PROTEINS IN AD AND MCI

Proteomics studies identified regionally specific HNE modification of proteins, i.e., ATP synthase, GS, MnSOD, and DRP-2 in AD hippocampus and

α -enolase, aconitase, aldolase, peroxiredoxin 6, and α -tubulin in AD cortex [75]. Some of these proteins were previously found to be either nitrated or carbonylated in AD [19, 33, 41–43, 45]. The appearance of different oxidative indices in target proteins modified by both protein carbonyls and 3-NT supports the role of oxidative stress in AD and is consistent with the notion that these specific proteins are highly vulnerable to oxidative modification and may be involved in AD.

In MCI hippocampus and cortex, increased levels of protein-bound HNE in neuropolypeptide h3, carbonyl reductase (NADPH), α -enolase, lactate dehydrogenase B, phosphoglycerate kinase, heat shock protein 70, ATP synthase α chain, pyruvate kinase, actin, elongation factor Tu, and translation initiation factor α were identified by proteomics [47]. Since most of the proteins that undergo HNE modification are dysfunctional, these proteomic results in amnesic MCI suggest that these HNE-bound proteins may be key players in the development of AD. Overlap of pathways involved in energy metabolism, neuritic abnormalities, cell cycle, tau phosphorylation, A β production, transcription and translation, mitochondrial abnormalities, and antioxidant system is consistent with the notion that these pathways could be involved in the progression of amnesic MCI to AD.

Increased lipid peroxidation in AD and MCI brain and a role for A β in this process were further supported by studies that showed loss of phospholipid asymmetry in AD and MCI brain, changes that are associated with apoptosis [76]. Noting that the high reactivity of free radicals requires that the initiator of lipid peroxidation must reside in the lipids, the findings above suggest that, in AD and MCI brain, oligomeric and hydrophobic A β inserts into the membrane of brain cells to cause lipid peroxidation and that such changes are an early event in the pathogenesis and progression of AD.

In conclusion, redox proteomics studies provided insights into key molecular pathways that, when oxidatively dysfunctional, play a significant role in the pathophysiology of AD [29]. Development of reliable and unique biomarkers to detect and diagnose MCI (or even earlier preclinical AD) and monitor drug efficacy in these disorders prior to cognitive decline will be of great importance for our rapidly aging population and may greatly accelerate the development of therapeutics and preventative approaches. In the future, redox proteomics of easily accessible bodily fluids or peripheral tissue in combination of other techniques, such as PET, MRI, and cognitive testing, may serve

as diagnostic tools to aid in monitoring the state of AD and therapeutic efficacy, and potentially provide a unique biomarker signature for AD. Other future studies in AD and MCI suggested by our redox proteomics results are therapeutic targeting (either pharmacologically or genetically) of key brain proteins that directly influence multiple AD-relevant pathways.

ACKNOWLEDGMENTS

Supported in part by the National Institutes of Health (AG05119 to DAB).

Authors' disclosures available online (<http://www.jalz.com/disclosures/view.php?id=1341>).

REFERENCES

- [1] Selkoe DJ (2001) Alzheimer's disease: Genes, proteins, and therapy. *Physiol Rev* **81**, 741-766.
- [2] Bertram L (2009) Alzheimer's disease genetics current status and future perspectives. *Int Rev Neurobiol* **84**, 167-184.
- [3] Bertram L, Tanzi RE (2009) Genome-wide association studies in Alzheimer's disease. *Hum Mol Genet* **18**, R137-R145.
- [4] Feulner TM, Laws SM, Friedrich P, Wagenpfeil S, Wurst SH, Riehle C, Kuhn KA, Krawczak M, Schreiber S, Nikolaus S, Forstl H, Kurz A, Riemenschneider M (2010) Examination of the current top candidate genes for AD in a genome-wide association study. *Mol Psychiatry* **15**, 756-766.
- [5] Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schurmann B, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Hull M, Rujescu D, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* **41**, 1088-1093.
- [6] Selkoe DJ (1996) Amyloid β -protein and the genetics of Alzheimer's disease. *J Biol Chem* **271**, 18295-18298.
- [7] Butterfield DA, Boyd-Kimball D (2005) The critical role of methionine 35 in Alzheimer's amyloid β -peptide (1-42)-induced oxidative stress and neurotoxicity. *Biochim Biophys Acta* **1703**, 149-156.
- [8] Butterfield DA, Sultana R (2011) Methionine-35 of $\text{A}\beta$ (1-42): Importance for oxidative stress in Alzheimer disease. *J Amino Acid* **2011**, 198430.
- [9] Butterfield DA, Bush AI (2004) Alzheimer's amyloid β -peptide (1-42): Involvement of methionine residue 35 in the oxidative stress and neurotoxicity properties of this peptide. *Neurobiol Aging* **25**, 563-568.
- [10] Sultana R, Butterfield DA (2004) Oxidatively modified GST and MRP1 in Alzheimer's disease brain: Implications for accumulation of reactive lipid peroxidation products. *Neurochem Res* **29**, 2215-2220.
- [11] Lauderback CM, Hackett JM, Huang FF, Keller JN, Szweda LI, Markesbery WR, Butterfield DA (2001) The glial glutamate transporter, GLT-1, is oxidatively modified by 4-hydroxy-2-nonenal in the Alzheimer's disease brain: The role of Abeta1-42. *J Neurochem* **78**, 413-416.
- [12] Butterfield DA, Hardas SS, Lange ML (2010) Oxidatively modified glyceraldehyde-3-phosphate dehydrogenase (GAPDH) and Alzheimer disease: Many pathways to neurodegeneration. *J Alzheimers Dis* **20**, 369-393.
- [13] Boyd-Kimball D, Sultana R, Mohammad-Abdul H, Butterfield DA (2005) Neurotoxicity and oxidative stress in D1M-substituted Alzheimer's A β (1-42): Relevance to N-terminal methionine chemistry in small model peptides. *Peptides* **26**, 665-673.
- [14] Schoneich C (2005) Methionine oxidation by reactive oxygen species: Reaction mechanisms and relevance to Alzheimer's disease. *Biochim Biophys Acta* **1703**, 111-119.
- [15] Jovanovic SV, Clements D, MacLeod K (1998) Biomarkers of oxidative stress are significantly elevated in Down syndrome. *Free Radic Biol Med* **25**, 1044-1048.
- [16] Cenini G, Dowling AL, Beckett TL, Barone E, Mancuso C, Murphy MP, Levine H 3rd, Lott IT, Schmitt FA, Butterfield DA, Head E (2012) Association between frontal cortex oxidative damage and beta-amyloid as a function of age in Down syndrome. *Biochim Biophys Acta* **1822**, 130-138.
- [17] Ansari MA, Scheff SW (2010) Oxidative stress in the progression of Alzheimer disease in the frontal cortex. *J Neuropathol Exp Neurol* **69**, 155-167.
- [18] Dalle-Donne I, Aldini G, Carini M, Colombo R, Rossi R, Milzani A (2006) Protein carbonylation, cellular dysfunction, and disease progression. *J Cell Mol Med* **10**, 389-406.
- [19] Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR, Butterfield DA (2006) Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* **22**, 76-87.
- [20] Smith MA, Richey Harris PL, Sayre LM, Beckman JS, Perry G (1997) Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J Neurosci* **17**, 2653-2657.
- [21] Markesbery WR, Kryscio RJ, Lovell MA, Morrow JD (2005) Lipid peroxidation is an early event in the brain in amnesic mild cognitive impairment. *Ann Neurol* **58**, 730-735.
- [22] Yao Y, Zhukareva V, Sung S, Clark CM, Rokach J, Lee VM, Trojanowski JQ, Pratico D (2003) Enhanced brain levels of 8,12-iso-iPF2 α -VI differentiate AD from frontotemporal dementia. *Neurology* **61**, 475-478.
- [23] Quinn JF, Montine KS, Moore M, Morrow JD, Kaye JA, Montine TJ (2004) Suppression of longitudinal increase in CSF F2-isoprostanes in Alzheimer's disease. *J Alzheimers Dis* **6**, 93-97.
- [24] Pratico D, Sung S (2004) Lipid peroxidation and oxidative imbalance: Early functional events in Alzheimer's disease. *J Alzheimers Dis* **6**, 171-175.
- [25] Pratico D, Clark CM, Liun F, Rokach J, Lee VY, Trojanowski JQ (2002) Increase of brain oxidative stress in mild cognitive impairment: A possible predictor of Alzheimer disease. *Arch Neurol* **59**, 972-976.
- [26] Keller JN, Schmitt FA, Scheff SW, Ding Q, Chen Q, Butterfield DA, Markesbery WR (2005) Evidence of increased

- oxidative damage in subjects with mild cognitive impairment. *Neurology* **64**, 1152-1156.
- [27] Butterfield DA, Perluigi M, Sultana R (2006) Oxidative stress in Alzheimer's disease brain: New insights from redox proteomics. *Eur J Pharmacol* **545**, 39-50.
- [28] Butterfield DA, Drake J, Pocernich C, Castegna A (2001) Evidence of oxidative damage in Alzheimer's disease brain: Central role for amyloid beta-peptide. *Trends Mol Med* **7**, 548-554.
- [29] Butterfield DA, Reed T, Newman SF, Sultana R (2007) Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. *Free Radic Biol Med* **43**, 658-677.
- [30] Markesbery WR (1997) Oxidative stress hypothesis in Alzheimer's disease. *Free Radic Biol Med* **23**, 134-147.
- [31] Nunomura A, Moreira PI, Lee HG, Zhu X, Castellani RJ, Smith MA, Perry G (2007) Neuronal death and survival under oxidative stress in Alzheimer and Parkinson diseases. *CNS Neurol Disord Drug Targets* **6**, 411-423.
- [32] Yatin SM, Varadarajan S, Link CD, Butterfield DA (1999) *In vitro* and *in vivo* oxidative stress associated with Alzheimer's amyloid beta-peptide (1-42). *Neurobiol Aging* **20**, 325-330; discussion 339-342.
- [33] Lyras L, Perry RH, Perry EK, Ince PG, Jenner A, Jenner P, Halliwell B (1998) Oxidative damage to proteins, lipids, and DNA in cortical brain regions from patients with dementia with Lewy bodies. *J Neurochem* **71**, 302-312.
- [34] Sultana R, Perluigi M, Butterfield DA (2006) Redox proteomics identification of oxidatively modified proteins in Alzheimer's disease brain and *in vivo* and *in vitro* models of AD centered around Aβ(1-42). *J Chromatogr B Analyt Technol Biomed Life Sci* **833**, 3-11.
- [35] Butterfield DA, Reed T, Perluigi M, De Marco C, Coccia R, Cini C, Sultana R (2006) Elevated protein-bound levels of the lipid peroxidation product, 4-hydroxy-2-nonenal, in brain from persons with mild cognitive impairment. *Neurosci Lett* **397**, 170-173.
- [36] Lovell MA, Markesbery WR (2008) Oxidatively modified RNA in mild cognitive impairment. *Neurobiol Dis* **29**, 169-175.
- [37] Aluise CD, Robinson RA, Cai J, Pierce WM, Markesbery WR, Butterfield DA (2011) Redox proteomics analysis of brains from subjects with amnesic mild cognitive impairment compared to brains from subjects with preclinical Alzheimer's disease: Insights into memory loss in MCI. *J Alzheimers Dis* **23**, 257-269.
- [38] Ebenezer PJ, Weidner AM, LeVine H 3rd, Markesbery WR, Murphy MP, Zhang L, Dasuri K, Fernandez-Kim SO, Bruce-Keller AJ, Gavilan E, Keller JN (2010) Neuron specific toxicity of oligomeric amyloid-beta: Role for JUN-kinase and oxidative stress. *J Alzheimers Dis* **22**, 839-848.
- [39] Butterfield DA, Gnjec A, Poon HF, Castegna A, Pierce WM, Klein JB, Martins RN (2006) Redox proteomics identification of oxidatively modified brain proteins in inherited Alzheimer's disease: An initial assessment. *J Alzheimers Dis* **10**, 391-397.
- [40] Butterfield A, Perluigi M, Reed T, Muharib T, Hughes CP, Robinson RA, Sultana R (2012) Redox proteomics in selected neurodegenerative disorders: From its infancy to future applications. *Antioxid Redox Signal*, doi:10.1089/ars.2011.4109.
- [41] Castegna A, Aksenov M, Aksenova M, Thongboonkerd V, Klein JB, Pierce WM, Booze R, Markesbery WR, Butterfield DA (2002) Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part I: Creatine kinase BB, glutamine synthase, and ubiquitin carboxy-terminal hydrolase L-1. *Free Radic Biol Med* **33**, 562-571.
- [42] Castegna A, Aksenov M, Thongboonkerd V, Klein JB, Pierce WM, Booze R, Markesbery WR, Butterfield DA (2002) Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part II: Dihydropyrimidinase-related protein 2, alpha-enolase and heat shock cognate 71. *J Neurochem* **82**, 1524-1532.
- [43] Castegna A, Thongboonkerd V, Klein JB, Lynn B, Markesbery WR, Butterfield DA (2003) Proteomic identification of nitrated proteins in Alzheimer's disease brain. *J Neurochem* **85**, 1394-1401.
- [44] Dalle-Donne I, Scaloni A, Butterfield DA (2006) *Redox Proteomics: From protein modifications to cellular dysfunction and diseases*. John Wiley and Sons, Hoboken, NJ.
- [45] Sultana R, Boyd-Kimball D, Poon HF, Cai J, Pierce WM, Klein JB, Merchant M, Markesbery WR, Butterfield DA (2006) Redox proteomics identification of oxidized proteins in Alzheimer's disease hippocampus and cerebellum: An approach to understand pathological and biochemical alterations in AD. *Neurobiol Aging* **27**, 1564-1576.
- [46] Sultana R, Reed T, Perluigi M, Coccia R, Pierce WM, Butterfield DA (2007) Proteomic identification of nitrated brain proteins in amnesic mild cognitive impairment: A regional study. *J Cell Mol Med* **11**, 839-851.
- [47] Reed T, Perluigi M, Sultana R, Pierce WM, Klein JB, Turner DM, Coccia R, Markesbery WR, Butterfield DA (2008) Redox proteomic identification of 4-hydroxy-2-nonenal-modified brain proteins in amnesic mild cognitive impairment: Insight into the role of lipid peroxidation in the progression and pathogenesis of Alzheimer's disease. *Neurobiol Dis* **30**, 107-120.
- [48] Aksenov MY, Aksenova MV, Butterfield DA, Geddes JW, Markesbery WR (2001) Protein oxidation in the brain in Alzheimer's disease. *Neuroscience* **103**, 373-383.
- [49] Choi J, Levey AI, Weintraub ST, Rees HD, Gearing M, Chin LS, Li L (2004) Oxidative modifications and down-regulation of ubiquitin carboxyl-terminal hydrolase L1 associated with idiopathic Parkinson's and Alzheimer's diseases. *J Biol Chem* **279**, 13256-13264.
- [50] Choi J, Forster MJ, McDonald SR, Weintraub ST, Carroll CA, Gracy RW (2004) Proteomic identification of specific oxidized proteins in ApoE-knockout mice: Relevance to Alzheimer's disease. *Free Radic Biol Med* **36**, 1155-1162.
- [51] Korolainen MA, Goldsteins G, Nyman TA, Alafuzoff I, Koistinaho J, Pirttila T (2006) Oxidative modification of proteins in the frontal cortex of Alzheimer's disease brain. *Neurobiol Aging* **27**, 42-53.
- [52] Choi J, Sullards MC, Olzmann JA, Rees HD, Weintraub ST, Bostwick DE, Gearing M, Levey AI, Chin LS, Li L (2006) Oxidative damage of DJ-1 is linked to sporadic Parkinson and Alzheimer diseases. *J Biol Chem* **281**, 10816-10824.
- [53] Choi J, Rees HD, Weintraub ST, Levey AI, Chin LS, Li L (2005) Oxidative modifications and aggregation of Cu, Zn-superoxide dismutase associated with Alzheimer and Parkinson diseases. *J Biol Chem* **280**, 11648-11655.
- [54] Terni B, Boada J, Portero-Otin M, Pamplona R, Ferrer I (2010) Mitochondrial ATP-Synthase in the Entorhinal Cortex Is a Target of Oxidative Stress at Stages I/II of Alzheimer's Disease Pathology. *Brain Pathol* **20**, 222-233.
- [55] Butterfield DA, Poon HF, St Clair D, Keller JN, Pierce WM, Klein JB, Markesbery WR (2006) Redox proteomics identification of oxidatively modified hippocampal proteins in mild cognitive impairment: Insights into the development of Alzheimer's disease. *Neurobiol Dis* **22**, 223-232.

- [56] Sultana R, Perluigi M, Butterfield DA (2009) Oxidatively modified proteins in Alzheimer's disease (AD), mild cognitive impairment and animal models of AD: Role of Abeta in pathogenesis. *Acta Neuropathol* **118**, 131-150.
- [57] Guix FX, Ill-Raga G, Bravo R, Nakaya T, de Fabritiis G, Coma M, Miscione GP, Villa-Freixa J, Suzuki T, Fernandez-Busquets X, Valverde MA, de Strooper B, Munoz FJ (2009) Amyloid-dependent triosephosphate isomerase nitrotyrosination induces glycation and tau fibrillation. *Brain* **132**, 1335-1345.
- [58] Reyes JF, Reynolds MR, Horowitz PM, Fu Y, Guillozet-Bongaarts AL, Berry R, Binder LI (2008) A possible link between astrocyte activation and tau nitration in Alzheimer's disease. *Neurobiol Dis* **31**, 198-208.
- [59] Butterfield DA, Reed TT, Perluigi M, De Marco C, Coccia R, Keller JN, Markesbery WR, Sultana R (2007) Elevated levels of 3-nitrotyrosine in brain from subjects with amnesic mild cognitive impairment: Implications for the role of nitration in the progression of Alzheimer's disease. *Brain Res* **1148**, 243-248.
- [60] Cenini G, Sultana R, Memo M, Butterfield DA (2008) Effects of oxidative and nitrosative stress in brain on p53 proapoptotic protein in amnesic mild cognitive impairment and Alzheimer disease. *Free Radic Biol Med* **45**, 81-85.
- [61] Riederer IM, Schiffrin M, Kovari E, Bouras C, Riederer BM (2009) Ubiquitination and cysteine nitrosylation during aging and Alzheimer's disease. *Brain Res Bull* **80**, 233-241.
- [62] Uehara T, Nakamura T, Yao D, Shi ZQ, Gu Z, Ma Y, Masliah E, Nomura Y, Lipton SA (2006) S-nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration. *Nature* **441**, 513-517.
- [63] Cho DH, Nakamura T, Fang J, Cieplak P, Godzik A, Gu Z, Lipton SA (2009) S-nitrosylation of Drp1 mediates beta-amyloid-related mitochondrial fission and neuronal injury. *Science* **324**, 102-105.
- [64] Martinez A, Portero-Otin M, Pamplona R, Ferrer I (2009) Protein targets of oxidative damage in human neurodegenerative diseases with abnormal protein aggregates. *Brain Pathol* **20**, 281-297.
- [65] Butterfield DA, Lange ML (2009) Multifunctional roles of enolase in Alzheimer's disease brain: Beyond altered glucose metabolism. *J Neurochem* **111**, 915-933.
- [66] Zheng L, Roeder RG, Luo Y (2003) S phase activation of the histone H2B promoter by OCA-S, a coactivator complex that contains GAPDH as a key component. *Cell* **114**, 255-266.
- [67] Bryksin AV, Laktionov PP (2008) Role of glyceraldehyde-3-phosphate dehydrogenase in vesicular transport from golgi apparatus to endoplasmic reticulum. *Biochemistry (Mosc)* **73**, 619-625.
- [68] Hara MR, Snyder SH (2006) Nitric oxide-GAPDH-Siah: A novel cell death cascade. *Cell Mol Neurobiol* **26**, 527-538.
- [69] Puder M, Soberman RJ (1997) Glutathione conjugates recognize the Rossmann fold of glyceraldehyde-3-phosphate dehydrogenase. *J Biol Chem* **272**, 10936-10940.
- [70] Schulze H, Schuler A, Stuber D, Dobeli H, Langen H, Huber G (1993) Rat brain glyceraldehyde-3-phosphate dehydrogenase interacts with the recombinant cytoplasmic domain of Alzheimer's beta-amyloid precursor protein. *J Neurochem* **60**, 1915-1922.
- [71] Pastorino L, Sun A, Lu PJ, Zhou XZ, Balastik M, Finn G, Wulf G, Lim J, Li SH, Li X, Xia W, Nicholson LK, Lu KP (2006) The prolyl isomerase Pin1 regulates amyloid precursor protein processing and amyloid-beta production. *Nature* **440**, 528-534.
- [72] Butterfield DA, Abdul HM, Opii W, Newman SF, Joshi G, Ansari MA, Sultana R (2006) Pin1 in Alzheimer's disease. *J Neurochem* **98**, 1697-1706.
- [73] Lu PJ, Wulf G, Zhou XZ, Davies P, Lu KP (1999) The prolyl isomerase Pin1 restores the function of Alzheimer-associated phosphorylated tau protein. *Nature* **399**, 784-788.
- [74] Sultana R, Butterfield DA (2010) Role of oxidative stress in the progression of Alzheimer's disease. *J Alzheimers Dis* **19**, 341-353.
- [75] Perluigi M, Sultana R, Cenini G, Di Domenico F, Memo M, Pierce WM, Coccia R, Butterfield DA (2009) Redox proteomics identification of HNE-modified brain proteins in Alzheimer's disease: Role of lipid peroxidation in Alzheimer's disease pathogenesis. *Proteomics Clin Appl* **13**, 682-693.
- [76] Bader Lange ML, Cenini G, Piroddi M, Abdul HM, Sultana R, Galli F, Memo M, Butterfield DA (2008) Loss of phospholipid asymmetry and elevated brain apoptotic protein levels in subjects with amnesic mild cognitive impairment and Alzheimer disease. *Neurobiol Dis* **29**, 456-464.